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Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation

Synonyms: LBSL, Mitochondrial Aspartyl-tRNA Synthetase Deficiency

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Summary

Clinical characteristics

Leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation (LBSL) is characterized by slowly progressive cerebellar ataxia and spasticity with dorsal column dysfunction (decreased position and vibration sense) in most individuals. The neurologic dysfunction involves the legs more than the arms. The tendon reflexes are retained. Deterioration of motor skills usually starts in childhood or adolescence, but occasionally not until adulthood. Dysarthria develops over time. Occasional findings include epilepsy; learning problems; cognitive decline; and reduced consciousness, neurologic deterioration, and fever following minor head trauma.

Individuals with neonatal or early-infantile onset have a severe disease course often associated with early death. Those with childhood onset have slow progression with wheelchair dependence in the teens or twenties. Adult onset is associated with slow progression and mild impairment.

Diagnosis/testing

The clinical diagnosis of LBSL can be established in a proband with characteristic abnormalities observed on brain and spinal cord MRI using MRI-based criteria. The molecular diagnosis can be established in a proband with suggestive findings and biallelic pathogenic variants in *DARS2* identified by molecular genetic testing. If molecular results are inconclusive, a functional assay to identify reduced MtAspRS enzyme activity in lymphoblasts can confirm the diagnosis.

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Management

Treatment of manifestations: Supportive therapy includes: physical therapy and rehabilitation to improve motor function and prevent contractures and scoliosis; and anti-seizure medication, speech therapy, special education, and social work support as needed.

Surveillance: Assess for new neurologic manifestations at each visit; monitor those with seizures as needed; consider brain MRI every few years to monitor progression; monitor developmental progress and educational needs at each visit throughout childhood; assess for family support needs at each visit.

Genetic counseling

LBSL is inherited in an autosomal recessive manner. If both parents are known to be heterozygous for a *DARS2* pathogenic variant, each sib of an affected individual has at conception a 25% chance of being affected, a 50% chance of being an asymptomatic carrier, and a 25% chance of being unaffected and not a carrier. Once the *DARS2* pathogenic variants have been identified in an affected family member, prenatal testing for a pregnancy at increased risk and preimplantation genetic testing are possible.

Diagnosis

Suggestive Findings

Diagnosis of leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation (LBSL) should be suspected in individuals with characteristic abnormalities observed on brain and spinal cord MRI [van der Knaap et al 2003, Scheper et al 2007, Steenweg et al 2012]. However, atypical presentations with antenatal or early-infantile onset characterized by either profound cerebral atrophy or a leukoencephalopathy in the absence of typical involvement of tracts in the brain stem and spinal cord have been described; see * NOTE: (1) [Stellingwerff et al 2021].

MRI Criteria for LBSL

See Steenweg et al [2012], Stellingwerff et al [2021], Figure 1, Figure 2, and Figure 3.

Major criteria. EITHER:

- Signal abnormalities (abnormally low signal on T₁-weighted images and abnormally high signal on T₂-weighted images) in the following:
 - Cerebral white matter, which is either nonhomogeneous and spotty or homogeneous and confluent, with relative sparing of the directly subcortical white matter
 - Dorsal columns and lateral corticospinal tracts of the spinal cord (Visualization of such abnormalities in the cervical spinal cord is sufficient.)
 - Pyramids and/or decussation of the medial lemniscus in the medulla oblongata

OR

- Profound atrophy of the cerebral hemispheres in infancy (often with numerous tortuous blood vessels at the brain surface in early stages)

Supportive criteria. Signal abnormalities in the following:

- Splenium of the corpus callosum
- Posterior limb of the internal capsule
- Superior cerebellar peduncles
- Inferior cerebellar peduncles

- Intraparenchymal part of the trigeminal nerve
- Mesencephalic trigeminal tracts
- Anterior spinocerebellar tracts in the medulla
- Cerebellar white matter

* NOTE: (1) Individuals with antenatal and early-infantile onset have either a cerebral cortical degeneration followed by profound cerebral atrophy or a leukoencephalopathy. The brain stem abnormalities typical of LBSL may or may not be present [Steenweg et al 2011, Stellingwerff et al 2021]. Therefore, individuals with early onset may not meet MRI criteria. (2) Lactate is elevated within the abnormal cerebral white matter in most but not all affected individuals [van der Knaap et al 2003, Steenweg et al 2011]. Lactate elevation in proton magnetic resonance spectroscopy of abnormal white matter has been mentioned as a diagnostic criterion but has a low distinguishing value. If the MRI meets the criteria for LBSL, this diagnosis should be considered, whether lactate is elevated or not. If the MRI does not meet the criteria for LBSL, elevated lactate may be a general indicator of a mitochondrial leukoencephalopathy, but not specifically LBSL.

Establishing the Diagnosis

Clinical Diagnosis

The clinical diagnosis of LBSL **can be established** in a proband with MRI-based criteria; all major criteria and at least one supportive criterion should be fulfilled (see MRI Criteria for LBSL) [Steenweg et al 2012]. See also Figure 1, Figure 2, and Figure 3.

Molecular Diagnosis

The molecular diagnosis of LBSL can be established in proband with suggestive findings and biallelic pathogenic variants in *DARS2* identified by molecular genetic testing (see Table 1).

If molecular results are inconclusive, a functional assay to identify reduced MtAspRS enzyme activity in lymphoblasts can confirm the diagnosis [van Berge et al 2014].

Molecular genetic testing approaches can include a combination of **gene-targeted testing** (single-gene testing, multigene panel) and **comprehensive genomic testing** (exome sequencing, genome sequencing, exome array) depending on the phenotype.

Gene-targeted testing requires that the clinician determine which gene(s) are likely involved, whereas genomic testing does not. Individuals with the distinctive findings described in Suggestive Findings are likely to be diagnosed using gene-targeted testing (see **Option 1**), whereas those with a phenotype indistinguishable from many other inherited disorders with leukoencephalopathy are more likely to be diagnosed using genomic testing (see **Option 2**).

Option 1

- **Single-gene testing.** Sequence analysis of *DARS2* is performed first to detect small intragenic deletions/insertions and missense, nonsense, and splice site variants. Note: Depending on the sequencing method used, single-exon, multiexon, or whole-gene deletions/duplications may not be detected. If only one or no variant is detected by the sequencing method used, the next step is to perform gene-targeted deletion/duplication analysis to detect exon and whole-gene deletions or duplications.
- **A multigene panel** that includes *DARS2* and other genes of interest (see Differential Diagnosis) may be considered to identify the genetic cause of the condition while limiting identification of variants of uncertain significance and pathogenic variants in genes that do not explain the underlying phenotype. Note: (1) The genes included in the panel and the diagnostic sensitivity of the testing used for each gene

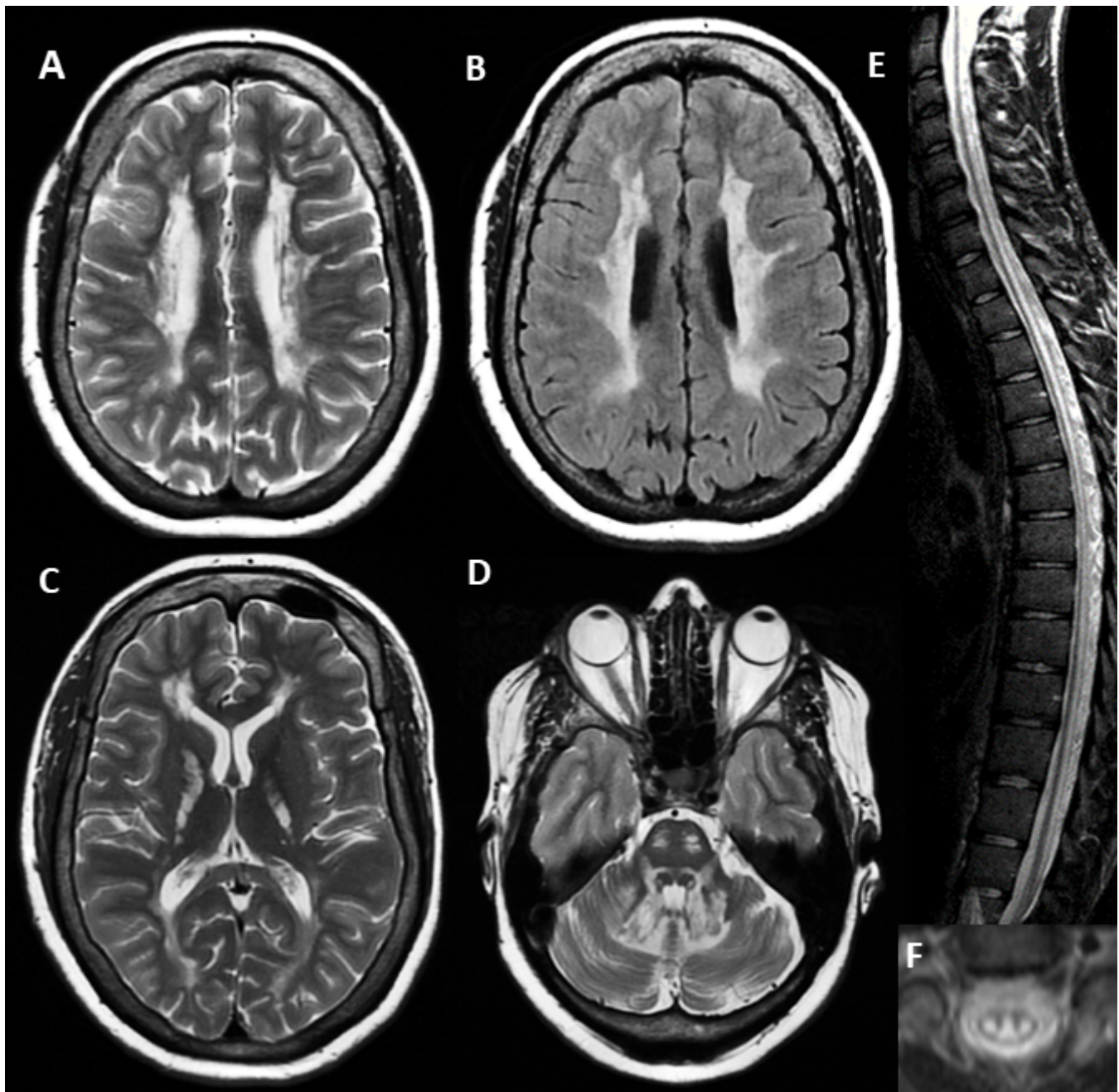


Figure 1. T₂-weighted (A, C, D) and FLAIR (B) axial images of the brain in an adult with LBSL show extensive, patchy abnormalities with increased signal in the periventricular and deep cerebral white matter and posterior limb of the internal capsule. In the posterior fossa the superior cerebellar peduncles, the intraparenchymal trajectories of the trigeminal nerves, the pyramidal tracts, and medial lemniscus in the pons are affected (D). T₂-weighted spinal cord images (E, F) show increased signal over the entire length of the spinal cord (E) with involvement of the (lateral) corticospinal tracts and dorsal columns (F).

vary by laboratory and are likely to change over time. (2) Some multigene panels may include genes not associated with the condition discussed in this *GeneReview*. (3) In some laboratories, panel options may include a custom laboratory-designed panel and/or custom phenotype-focused exome analysis that includes genes specified by the clinician. (4) Methods used in a panel may include sequence analysis, deletion/duplication analysis, and/or other non-sequencing-based tests.

For an introduction to multigene panels click [here](#). More detailed information for clinicians ordering genetic tests can be found [here](#).

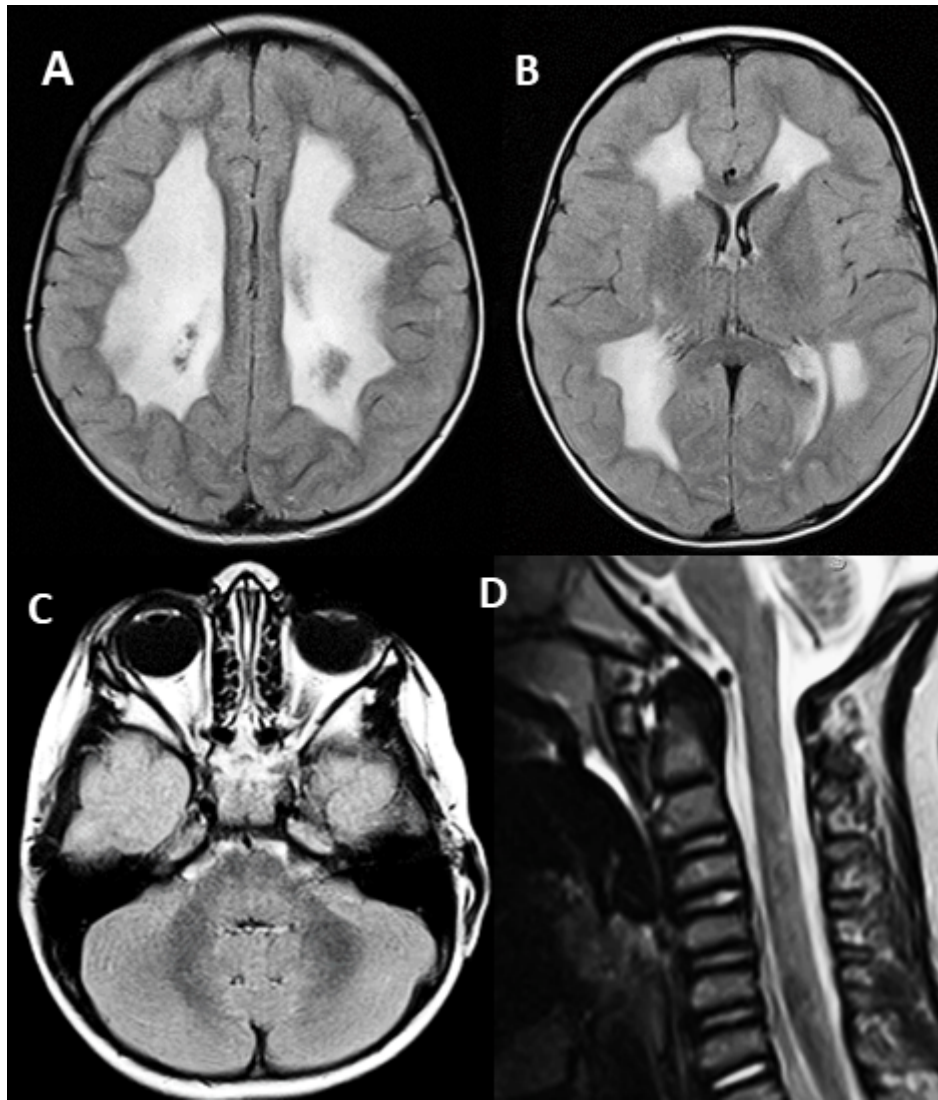


Figure 2. Axial FLAIR images of the brain (A-C) in a young individual (toddler) with early presentation of LBSL show abnormal signal intensity (with some areas of rarefaction) of almost all cerebral white matter, only sparing the directly subcortical areas. The internal capsule (B), brain stem (C), cerebellum (C), and spinal cord (D) are spared.

Images for this figure courtesy of Dr Menno Stellingwerff

Option 2. When the phenotype is indistinguishable from many other inherited disorders characterized by leukoencephalopathy, **comprehensive genomic testing** (which does not require the clinician to determine which gene is likely involved) is an option. **Exome sequencing** is most commonly used; **genome sequencing** is also possible. Importantly, the most common *DARS2* pathogenic variant involves intron 2, which is easily missed by exome sequencing. If a heterozygous *DARS2* variant is identified in an individual with leukoencephalopathy, analysis of intron 2 should be performed.

If exome sequencing is not diagnostic, **exome array** (when clinically available) may be considered to detect (multi)exon deletions or duplications that cannot be detected by sequence analysis [Lan et al 2017].

For an introduction to comprehensive genomic testing click [here](#). More detailed information for clinicians ordering genomic testing can be found [here](#).

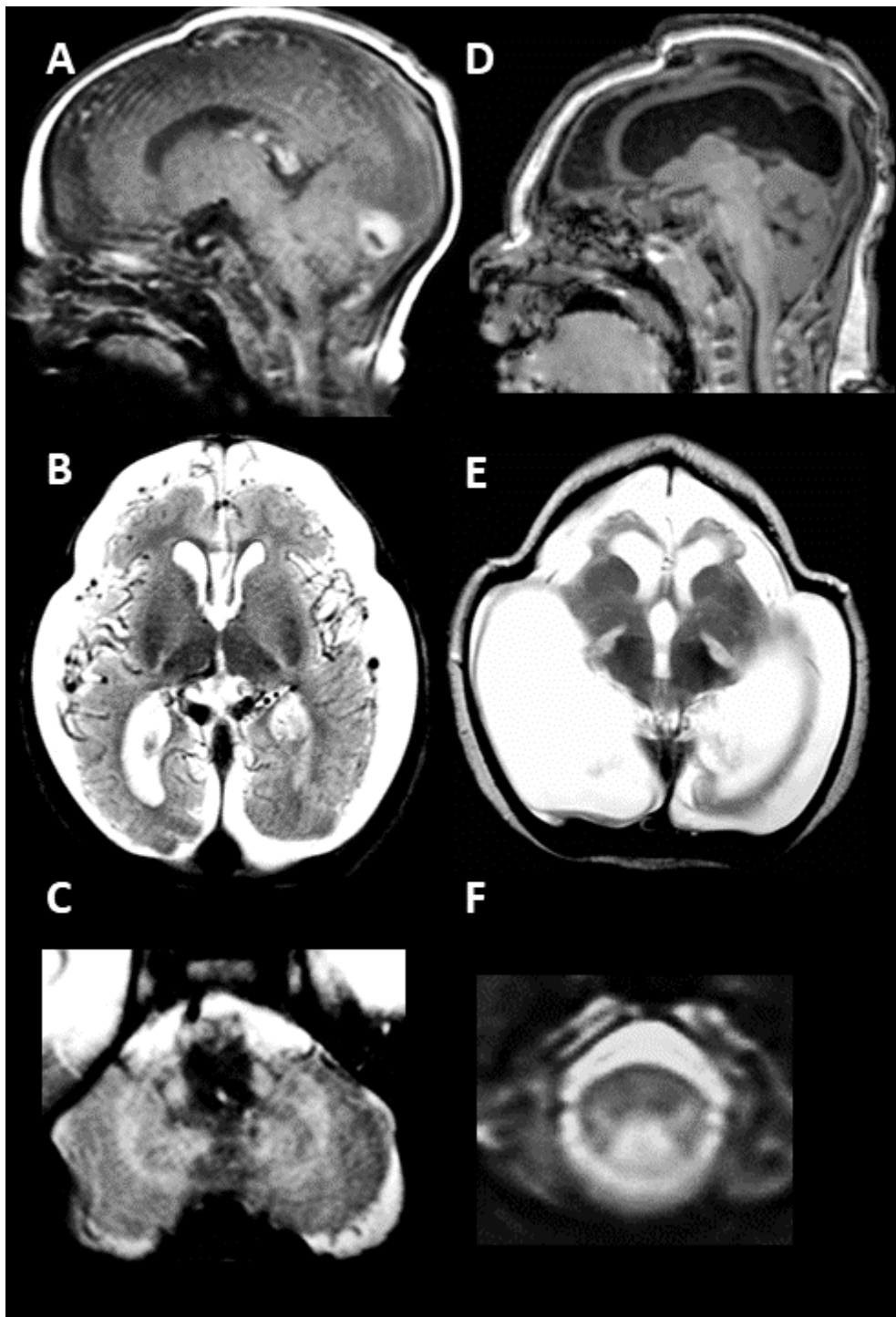


Figure 3. The MRI in a severe, early-onset form of LBSL at age two days (A, B, C) and age five months (D, E, F) shows striking progression of abnormalities. At two days, sagittal T₁-weighted (A) and axial T₂-weighted images (B) show hypoplasia of the cerebrum with numerous tortuous blood vessels on the surface of the brain (A, B). In the medulla, the inferior cerebellar peduncles and pyramids are clearly affected (C). At follow up, the cerebral mantle has been reduced to a thin rim, with degeneration of virtually all cortex and white matter (D, E). The tortuous vessels are no longer present. The posterior limb of the internal capsule is abnormal in signal (E). The axial image through the cervical spinal cord (F) shows signal abnormalities in the corticospinal tracts and dorsal columns.

Images for this figure courtesy of Dr Menno Stellingwerff

Table 1. Molecular Genetic Testing Used in Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation

Gene ¹	Method	Proportion of Pathogenic Variants ² Detectable by This Method
<i>DARS2</i>	Sequence analysis ³	~90% ^{4, 5}
	Gene-targeted deletion/duplication analysis ⁶	Rare ⁷
Unknown	NA	See footnote 8.

1. See Table A. Genes and Databases for chromosome locus and protein.

2. See Molecular Genetics for information on allelic variants detected in this gene.

3. Sequence analysis detects variants that are benign, likely benign, of unknown significance, likely pathogenic, or pathogenic. Variants may include small intragenic deletions/insertions and missense, nonsense, and splice site variants; typically, exon or whole-gene deletions/duplications are not detected. For issues to consider in interpretation of sequence analysis results, click [here](#).

4. Affected individuals are almost invariably compound heterozygous for two pathogenic variants in *DARS2*. A few individuals with homozygous pathogenic variants have been identified.

5. In a few individuals it has not been possible to determine both pathogenic variants: in four of 43 families the second pathogenic variant could not be found in gDNA; in two of three the second pathogenic variant was detected using cDNA; cells of the fourth individual were not available for isolation of mRNA for cDNA synthesis.

6. Gene-targeted deletion/duplication analysis detects intragenic deletions or duplications. Methods used may include quantitative PCR, long-range PCR, multiplex ligation-dependent probe amplification (MLPA), and a gene-targeted microarray designed to detect single-exon deletions or duplications.

7. Intragenic deletion of exon 12 has been reported [Lan et al 2017].

8. In one person fulfilling the MRI criteria for LBSL, no pathogenic variants in *DARS2* were detected in either gDNA or cDNA [Scheper et al 2007; Scheper & van der Knaap, personal communication].

Clinical Characteristics

Clinical Description

The clinical picture of LBSL consists of slowly progressive cerebellar ataxia, spasticity, and dorsal column dysfunction, involving the legs more than the arms. Most affected individuals have decreased position and vibration sense of the legs more than the arms, leading to increased difficulty walking in the dark. Manual dexterity becomes impaired to a variable degree.

To date, more than 100 individuals have been identified with biallelic pathogenic variants in *DARS2*. Large series [Steenweg et al 2011, van Berge et al 2014, Stellingwerff et al 2021] as well as multiple case reports [Köhler et al 2015, Shimojima et al 2017, Yahia et al 2018, Yelam et al 2019, N'Gbo N'Gbo Ikazabo et al 2020, Yazici Gencdal et al 2020] have been described. Prospective natural history data are currently not available. The following description of the phenotypic features associated with this condition is based on the cited reports.

Table 2. Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation: Frequency of Select Features

Feature	% of Persons w/Feature	Comment
Normal early development	80%	Infantile form was recently identified [Stellingwerff et al 2021]; relative incidence is not clear yet but probably low.
Loss of motor skills	100%	Core feature of LBSL
Cerebellar ataxia	Very common	
Spasticity	Very common	
Axonal neuropathy	Uncommon	↓ or absent tendon reflexes, distal weakness, sensory loss
Learning disability	20%	Most often learning disability; severe intellectual impairment is rare.

Table 2. continued from previous page.

Feature	% of Persons w/Feature	Comment
Cognitive decline	20%	
Epilepsy	Uncommon	Usually easily controlled w/ASM

ASM = anti-seizure medication

Variable severity. The range of disease spectrum includes:

- **Antenatal and early-infantile onset.** Severe form characterized by microcephaly, severe developmental delay, epilepsy, and often early death [Steenweg et al 2012, Stellingwerff et al 2021]
- **Childhood onset.** Most common form with slow progression. Most individuals with childhood onset become partially or fully wheelchair dependent in their teens, twenties, or later [van Berge et al 2014].
- **Adult onset.** Slowly progressive form with only mild impairment [van Berge et al 2014]. Individuals with adult-onset disease are not known to become wheelchair dependent [van Berge et al 2014].

Motor involvement. In most affected individuals, initial motor development is normal. Deterioration of motor skills usually starts in childhood or adolescence [Uluc et al 2008, van Berge et al 2014] and occasionally in infancy [Steenweg et al 2012] or adulthood [van Berge et al 2014]. Slowly progressive cerebellar ataxia and spasticity occur in most individuals. Axonal peripheral neuropathy may add to the distal weakness. The motor dysfunction involves the legs more than the arms.

In the individuals with antenatal or early-infantile onset, motor involvement is severe and typically no or very few motor milestones are achieved [Stellingwerff et al 2021].

Sensory involvement. There is severe dorsal column dysfunction with decreased position and vibration sense on neurologic examination, causing sensory ataxia especially in the legs. Evidence of an axonal neuropathy is found in some (not all) affected individuals, leading to decreased or absent tendon reflexes and distal sensory loss affecting the legs more than the arms [van der Knaap et al 2003, Távora et al 2007, Uluc et al 2008, Isohanni et al 2010]. However, true peripheral neuropathy is probably not common as neurophysiologic examination is often normal [van der Knaap et al 2003].

Speech. Dysarthria develops over time. Swallowing is generally not affected in individuals with childhood or adult onset [van Berge et al 2014].

Cognitive skills. Some have learning problems from early on, but most have normal intellectual capacity. Cognitive decline may occur and is usually slow [van der Knaap et al 2003, Serkov et al 2004]. Individuals with antenatal or early-infantile presentation typically have severe cognitive impairment [Stellingwerff et al 2021].

Epilepsy. Some affected individuals develop epilepsy. Seizures are infrequent and easily controlled with medication [van der Knaap et al 2003]. In individuals with antenatal and early-infantile onset, epilepsy is almost invariably present and often severe [Stellingwerff et al 2021].

Microcephaly. In individuals with antenatal or early-infantile onset, microcephaly is almost invariably present and often severe [Stellingwerff et al 2021]. See also Figure 3.

Response to minor head trauma. Some affected individuals experience lowered consciousness, neurologic deterioration, and fever following minor head trauma [Serkov et al 2004]. Recovery is only partial.

Routine laboratory tests, including cerebrospinal fluid (CSF) analysis, are usually normal. In a few individuals mild and inconsistent mild elevation of lactate concentration has been noted in blood or CSF or both. No published information is available.

Neuropathologic findings have been described in two sibs with a severe variant of LBSL [Yamashita et al 2013]. Electron microscopy revealed vacuolar changes and myelin splitting in the affected white matter [Yamashita et al 2013]. Quantitative MR parameters are in line with these findings, with T₂ hyperintense white matter showing increased fractional anisotropy and reduced apparent diffusion coefficient [Steenweg et al 2011].

Genotype-Phenotype Correlations

An overview study of 66 affected individuals revealed preliminary evidence in support of a genotype-phenotype correlation [van Berge et al 2014]. A recent study on antenatal and early-infantile presentation of LBSL substantiates genotype-phenotype correlation. Individuals with this presentation lack the common "leaky" splice site variants and instead have more severe loss-of-function variants [Stellingwerff et al 2021] (see Molecular Pathogenesis).

Prevalence

LBSL is rare. Carrier rate in the general population is low, with the exception of Finland, where a carrier rate of 1:95 has been reported [Isohanni et al 2010]. To date, only one family with parental consanguinity has been observed [Miyake et al 2011]. Almost all affected individuals are compound heterozygous for two pathogenic variants. Only four individuals from two families have been described with homozygous pathogenic variants [Miyake et al 2011, Synofzik et al 2011]. Strikingly, no instances of homozygosity have been seen among Finnish persons with LBSL [van Berge et al 2014].

Genetically Related (Allelic) Disorders

No phenotypes other than those discussed in this *GeneReview* are known to be associated with germline pathogenic variants in *DARS2*.

Differential Diagnosis

Classic Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation

The classic clinical picture of leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation (LBSL) consists of slowly progressive cerebellar ataxia, spasticity, and dorsal column dysfunction, involving the legs more than the arms. The tendon reflexes are retained. Based on these findings alone, many disorders can be considered [Parodi et al 2018]; however, characteristic brain and spinal cord MRI findings (see Figure 1) distinguish LBSL from other disorders with overlapping clinical features.

Ataxia. MRI findings distinguish LBSL from other [hereditary ataxias](#) [Steenweg et al 2011, van der Knaap et al 2019]. Specifically, the spotty white matter abnormalities with focal diffusion restriction seen in [CSF1R-related adult-onset leukoencephalopathy with axonal spheroids and pigmented glia \(ALSP\)](#), an autosomal dominant disorder, can be confused with adult LBSL; however, in ALSP, the brain stem and spinal cord involvement typical of LBSL are lacking [Steenweg et al 2011, Mochel et al 2019].

Ataxia and spasticity in combination with MRI abnormalities

- The clinical findings of ataxia and spasticity in combination with MRI abnormalities of the dorsal columns, lateral corticospinal tracts, and cerebral white matter are compatible with vitamin B₁₂ deficiency (combined cord degeneration) [Locatelli et al 1999]. However, the brain stem abnormalities typically seen in LBSL do not occur in vitamin B₁₂ deficiency. In vitamin B₁₂ deficiency the cervical spinal cord is mainly affected [Locatelli et al 1999], whereas in LBSL the entire spinal cord is affected [van der Knaap et al 2003]. See also Figure 1.

- Hypomyelination with brain stem and spinal cord involvement and leg spasticity (HBSL; OMIM 615281), caused by biallelic pathogenic variants in *DARS1*, shares part of the selective vulnerability of brain stem and spinal cord structures, but the signal abnormalities in HBSL are mild – indicative of hypomyelination – while they are prominent in LBSL [Taft et al 2013]. See also Figure 1 and Figure 2.

Elevated lactate in MRS or body fluids or both combined with clinical findings of a spinocerebellar ataxia or white matter abnormalities on MRI or both should lead to the consideration of [mitochondrial disorders](#) [Vernon & Bindoff 2018]. Although the brain stem and spinal cord are frequently affected in mitochondrial disorders, the selective involvement of specific brain stem and spinal cord tracts is unique for LBSL [van der Knaap & Valk 2005].

Variant LBSL: Antenatal and Early-Infantile Onset

A variant form of LBSL is characterized by antenatal or early infantile presentation and profoundly delayed or absent development and epilepsy [Stellingwerff et al 2021]. MRI reveals severe cerebral atrophy. If the brain stem and spinal cord abnormalities typical of LBSL are present, the correct diagnosis can still be suggested. However, in some individuals with early-onset LBSL, brain stem and spinal cord abnormalities are absent and in such cases the MRI shows a picture of an early onset severe neuronal degenerative disorder (see Figure 3). In these individuals, LBSL needs to be distinguished from infantile neuronal ceroid lipofuscinosis (OMIM PS256730), and other early-onset tRNA synthetase defects related to variants in *AARS1* (OMIM 616339) and *RARS1* (OMIM 616140) [Simons et al 2015, Mendes et al 2020].

Additionally, Stellingwerff et al [2021] reported an infantile-onset LBSL variant form with a leukoencephalopathy but lacking the typical brain stem or spinal cord abnormalities (see Figure 2). In such cases other mitochondrial leukodystrophies should be considered [Zhang et al 2019].

Management

Evaluations Following Initial Diagnosis

To establish the extent of disease and needs in an individual diagnosed with leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation (LBSL), the evaluations summarized in Table 3 (if not performed as part of the evaluation that led to the diagnosis) are recommended.

Table 3. Recommended Evaluations Following Initial Diagnosis in Individuals with Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation

System/Concern	Evaluation	Comment
Neurologic/ Neurodevelopment	Neurologic exam	Consider: <ul style="list-style-type: none"> • Brain & spinal cord MRI & proton MRS of abnormal cerebral white matter; • EEG if seizures are suspected.
	PT/OT/rehab medicine assessment	To assess for weakness, spasticity, & gait issues
	Speech therapy assessment	In those w/dysarthria
	Cognitive assessment	In those w/suspected cognitive involvement
Genetic counseling	By genetics professionals ¹	To inform patients & families re nature, MOI, & implications of LBSL to facilitate medical & personal decision making

Table 3. continued from previous page.

System/Concern	Evaluation	Comment
Family support/resources	For affected infants/children assess: <ul style="list-style-type: none"> • Use of community or online resources such as Parent to Parent; • Need for social work involvement for parental support; • Need for home nursing referral. 	

LBSL = leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation; MOI = mode of inheritance; OT = occupational therapy; PT = physical therapy

1. Medical geneticist, certified genetic counselor, or certified advanced genetic nurse

Treatment of Manifestations

Table 4. Treatment of Manifestations in Individuals with Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation

Manifestation/Concern	Treatment	Considerations/Other
Spasticity	PT & rehab medicine to improve motor function & prevent contractures & scoliosis	Assess need for positioning & mobility devices, disability parking placard.
Seizures	ASM if epileptic seizures are present	
Dysarthria	Speech therapy	
Intellectual disability	Early intervention & IEP as indicated	
Family/Community	<ul style="list-style-type: none"> • Ensure appropriate social work involvement to connect families w/local resources, respite, & support. • Care coordination to manage multiple subspecialty appointments, equipment, medications, & supplies 	Ongoing assessment of need for palliative care involvement &/or home nursing

ASM = anti-seizure medication; IEP = individualized education plan; PT = physical therapy

Surveillance

Table 5. Recommended Surveillance for Individuals with Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation

System/Concern	Evaluation	Frequency
Neurologic	Assess for new manifestations (e.g., seizures, changes in tone, movement disorders).	At each visit
	Monitor those w/seizures.	As clinically indicated
	Brain MRI	Can be considered every few yrs to monitor progression
Development	Monitor developmental progress & educational needs.	At each visit throughout childhood
Miscellaneous/Other	Assess family need for social work support (e.g., palliative/respite care, home nursing; other local resources) & care coordination.	At each visit

Agents/Circumstances to Avoid

Case reports have described neurologic deterioration following head trauma (as has been reported for several other leukodystrophies including vanishing white matter and adrenoleukodystrophy). The exact risk is not clear. No specific guidelines exist for contact sports or other activities. Affected individuals should be counseled on the

above-mentioned observations and can then make an individual decision based on personal preference [van der Knaap et al 2006, Budhram & Pandey 2017].

Evaluation of Relatives at Risk

See Genetic Counseling for issues related to testing of at-risk relatives for genetic counseling purposes.

Therapies Under Investigation

Search [ClinicalTrials.gov](https://clinicaltrials.gov) in the US and [EU Clinical Trials Register](https://clinicaltrialsregister.eu) in Europe for access to information on clinical studies for a wide range of diseases and conditions. Note: There may not be clinical trials for this disorder.

Other

Studies of muscle biopsies, fibroblasts, and lymphoblasts show no evidence of mitochondrial dysfunction; therefore, there is no rationale for the "mitochondrial cocktail" of vitamins and cofactors, often given to persons with mitochondrial dysfunction.

Genetic Counseling

Genetic counseling is the process of providing individuals and families with information on the nature, mode(s) of inheritance, and implications of genetic disorders to help them make informed medical and personal decisions. The following section deals with genetic risk assessment and the use of family history and genetic testing to clarify genetic status for family members; it is not meant to address all personal, cultural, or ethical issues that may arise or to substitute for consultation with a genetics professional. —ED.

Mode of Inheritance

Leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation (LBSL) is inherited in an autosomal recessive manner.

Risk to Family Members

Parents of a proband

- The parents of an affected individual are obligate heterozygotes (i.e., presumed to be carriers of one *DARS2* pathogenic variant based on family history).
- Molecular genetic testing is recommended for the parents of a proband to confirm that both parents are heterozygous for a *DARS2* pathogenic variant and to allow reliable recurrence risk assessment. If a pathogenic variant is detected in only one parent, the following possibilities should be considered:
 - One of the pathogenic variants identified in the proband occurred as a *de novo* event in the proband or as a postzygotic *de novo* event in a mosaic parent [Jónsson et al 2017].
 - Uniparental isodisomy for the parental chromosome with the pathogenic variant resulted in homozygosity for the pathogenic variant in the proband.
- Heterozygotes (carriers) are asymptomatic and are not at risk of developing the disorder.

Sibs of a proband

- If both parents are known to be heterozygous for a *DARS2* pathogenic variant, each sib of an affected individual has at conception a 25% chance of being affected, a 50% chance of being an asymptomatic carrier, and a 25% chance of being unaffected and not a carrier.

- The clinical manifestations in sibs with biallelic *DARS2* pathogenic variant are highly variable. Yahia et al [2018] described clinical presentations ranging "from an apparently healthy individual (with findings limited to brisk reflexes incidentally recognized at age 20 years) to disabled patients" within the same family.
- Heterozygotes (carriers) are asymptomatic and are not at risk of developing the disorder.

Offspring of a proband. The offspring of an individual with LBSL are obligate heterozygotes (carriers) for a pathogenic variant in *DARS2*.

Other family members. Each sib of the proband's parents is at a 50% risk of being a carrier of a *DARS2* pathogenic variant.

Carrier Detection

Carrier testing for at-risk family members requires prior identification of the *DARS2* pathogenic variants in the family.

Related Genetic Counseling Issues

Family planning

- The optimal time for determination of genetic risk and discussion of the availability of prenatal/preimplantation genetic testing is before pregnancy.
- It is appropriate to offer genetic counseling (including discussion of potential risks to offspring and reproductive options) to young adults who are affected, are carriers, or are at risk of being carriers.

DNA banking. Because it is likely that testing methodology and our understanding of genes, allelic variants, and diseases will improve in the future, consideration should be given to banking DNA from probands in whom a molecular diagnosis has not been confirmed (i.e., the causative genetic alteration/s are unknown).

Prenatal Testing and Preimplantation Genetic Testing

Once the *DARS2* pathogenic variants have been identified in an affected family member, prenatal testing for a pregnancy at increased risk and preimplantation genetic testing are possible.

Differences in perspective may exist among medical professionals and within families regarding the use of prenatal testing. While most centers would consider use of prenatal testing to be a personal decision, discussion of these issues may be helpful.

Resources

GeneReviews staff has selected the following disease-specific and/or umbrella support organizations and/or registries for the benefit of individuals with this disorder and their families. GeneReviews is not responsible for the information provided by other organizations. For information on selection criteria, click [here](#).

- **A Cure for Ellie / Cure LBSL**
Email: ACureforEllie@gmail.com
www.curelbls.org
- **National Organization for Rare Disorders (NORD)**
[Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation](#)
- **European Leukodystrophy Association (ELA)**
Phone: 03 83 30 93 34

www.ela-asso.com

- **EURORDIS-Rare Diseases Europe**
Email: eurordis@eurordis.org
[Find a patient organization](#)
- **Metabolic Support UK**
 United Kingdom
Phone: 0845 241 2173
metabolicsupportuk.org
- **United Leukodystrophy Foundation**
Phone: 800-SAV-LIVE; 815-748-3211
Email: office@ulf.org
www.ulf.org
- **Myelin Disorders Bioregistry Project**
Phone: 215-590-1719
Email: sherbinio@chop.edu
[Myelin Disorders Bioregistry Project](#)

Molecular Genetics

Information in the Molecular Genetics and OMIM tables may differ from that elsewhere in the GeneReview: tables may contain more recent information. —ED.

Table A. Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation: Genes and Databases

Gene	Chromosome Locus	Protein	Locus-Specific Databases	HGMD	ClinVar
DARS2	1q25.1	Aspartate--tRNA ligase, mitochondrial	DARS2 database	DARS2	DARS2

Data are compiled from the following standard references: gene from [HGNC](#); chromosome locus from [OMIM](#); protein from [UniProt](#). For a description of databases (Locus Specific, HGMD, ClinVar) to which links are provided, click [here](#).

Table B. OMIM Entries for Leukoencephalopathy with Brain Stem and Spinal Cord Involvement and Lactate Elevation ([View All in OMIM](#))

610956	ASPARTYL-tRNA SYNTHETASE 2; DARS2
611105	LEUKOENCEPHALOPATHY WITH BRAINSTEM AND SPINAL CORD INVOLVEMENT AND LACTATE ELEVATION; LBSL

Molecular Pathogenesis

DARS2 encodes a mitochondrial aspartyl-tRNA synthetase (mtAspRS), which charges tRNA^{Asp} in mitochondria. Mitochondrial AspRS possesses strictly conserved residues found in all known AspRS sequences, including those for ATP binding and tRNA binding. Residues involved in amino acid binding include those typical for class II aaRSs and those specific for aspartic acid recognition [Bonfond et al 2005]. Based on the homology to its bacterial counterparts, mtAspRS is thought to form homodimers [Delarue et al 1994].

Almost all affected individuals have a pathogenic variant that affects splicing of exon 3. Incorrect splicing of this exon results in a frameshift in the reading frame and nonsense-mediated decay of the wrongly spliced mRNA. It should be noted that these pathogenic variants upstream of exon 3 diminish but do not completely abolish

correct splicing. As a result, a low amount of wild type protein is produced in the cells of an affected individual. A total lack of mtAspRS activity is thought to be incompatible with life.

Another common pathogenic variant, c.492+2T>C (p.Met134_Lys165del) leads to a deletion of part of the protein. Though not fully understood, this deletion is likely to have a severe effect on the function of the protein.

Mechanism of disease causation. Loss of function. Several pathogenic missense variants have been shown to severely reduce the amino acylation function in assays with purified bacterially expressed recombinant proteins [Scheper et al 2007].

DARS2-specific laboratory technical considerations. In almost all affected individuals one pathogenic variant is present upstream of exon 3. The variant c.228-21_228-20delTTinsC is most often observed. In other affected individuals nucleotide changes are seen in the same region, within a stretch of ten or 11 C-residues that lies ten nucleotides upstream of exon 3 [Scheper et al 2007].

Table 6. Notable *DARS2* Pathogenic Variants

Reference Sequences	DNA Nucleotide Change	Predicted Protein Change	Comment [Reference]
NM_108122.5 NP_060592.2	c.228-21_228-20delTTinsC	p.Arg76SerfsTer5	Most common pathogenic variant reported
	c.492+2T>C	p.Met134_Lys165del	See footnote 1.
	c.455G>T	p.Cys152Phe	
NG_016138.1	Exon 12 deletion ²	See footnote 2.	Lan et al [2017]

Variants listed in the table have been provided by the authors. *GeneReviews* staff have not independently verified the classification of variants.

GeneReviews follows the standard naming conventions of the Human Genome Variation Society (varnomen.hgvs.org). See [Quick Reference](#) for an explanation of nomenclature.

1. Several individuals share haplotypes involving five or six microsatellite markers on chromosome 1p25. The pathogenic variants c.492+2T>C and c.455G>T are correlated with two of these haplotypes and are often seen in affected individuals of northeastern European origin [Scheper et al 2007, Isohanni et al 2010].

2. Variant designation that does not conform to current naming conventions; no breakpoints were delineated in the original report.

Chapter Notes

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References

Literature Cited

- Bonnefond L, Fender A, Rudinger-Thirion J, Giegé R, Florentz C, Sissler M. Toward the full set of human mitochondrial aminoacyl-tRNA synthetases: characterization of AspRS and TyrRS. *Biochemistry*. 2005;44:4805–16. PubMed PMID: 15779907.
- Budhram A, Pandey SK. Activation of cerebral X-linked adrenoleukodystrophy after head trauma. *Can J Neurol Sci*. 2017;44:597–8. PubMed PMID: 28534457.
- Delarue M, Poterszman A, Nikonov S, Garber M, Moras D, Thierry JC. Crystal structure of a prokaryotic aspartyl tRNA-synthetase. *EMBO J*. 1994;13:3219–29. PubMed PMID: 8045252.
- Isohanni P, Linnankivi T, Buzkova J, Lonqvist T, Pihko H, Valanne L, Tienari PJ, Elovaara I, Pirttila T, Reunanen M, Koivisto K, Marjavaara S, Suomalainen A. DARS2 mutations in mitochondrial leukoencephalopathy and multiple sclerosis. *J Med Genet*. 2010;47:66–70. PubMed PMID: 19592391.
- Jónsson H, Sulem P, Kehr B, Kristmundsdottir S, Zink F, Hjartarson E, Hardarson MT, Hjorleifsson KE, Eggertsson HP, Gudjonsson SA, Ward LD, Arnadottir GA, Helgason EA, Helgason H, Gylfason A, Jonasdottir A, Jonasdottir A, Rafnar T, Frigge M, Stacey SN, Th Magnusson O, Thorsteinsdottir U, Masson G, Kong A, Halldorsson BV, Helgason A, Gudbjartsson DF, Stefansson K. Parental influence on human germline de novo mutations in 1,548 trios from Iceland. *Nature*. 2017;549:519–22. PubMed PMID: 28959963.
- Köhler C, Heyer C, Hoffjan S, Stemmler S, Lücke T, Thielsa C, Kohlschütter A, Lobe U, Horvath R, Kleinle S, Benet-Pages A, Abicht A. Early-onset leukoencephalopathy due to a homozygous missense mutation in the DARS2 gene. *Mol Cell Probes*. 2015;29:319–22. PubMed PMID: 26327357.
- Lan MY, Chang YY, Yeh TH, Lin TK, Lu CS. Leukoencephalopathy with brainstem and spinal cord involvement and lactate elevation (LBSL) with a novel DARS2 mutation and isolated progressive spastic paraparesis. *J Neurol Sci*. 2017;372:229–31. PubMed PMID: 28017220.
- Locatelli ER, Lauren R, Ballard P, Mark AS. MRI in vitamin B12 deficiency myelopathy. *Can J Neurol Sci*. 1999;26:60–3. PubMed PMID: 10068811.
- Mendes MI, Green LMC, Bertini E, Tonduti D, Aiello C, Smith D, Salsano E, Beerepoot S, Hertecant J, von Spiczak S, Livingston JH, Emrick L, Fraser J, Russell L, Bernard G, Magri S, Di Bella D, Taroni F, Koenig MK, Moroni I, Cappuccio G, Brunetti-Pierri N, Rhee J, Mendelsohn BA, Helbig I, Helbig K, Muhle H, Ismayl O, Vanderver AL, Salomons GS, van der Knaap MS, Wolf NI. RARS1-related hypomyelinating leukodystrophy: expanding the spectrum. *Ann Clin Transl Neurol*. 2020;7:83–93. PubMed PMID: 31814314.
- Miyake N, Yamashita S, Kurosawa K, Miyatake S, Tsurusaki Y, Doi H, Saito H, Matsumoto N. A novel homozygous mutation of DARS2 may cause a severe LBSL variant. *Clin Genet*. 2011;80:293–6. PubMed PMID: 21815884.
- Mochel F, Delorme C, Czernecki V, Froger J, Cormier F, Ellie E, Fegueux N, Lehericy S, Lumbroso S, Schiffmann R, Aubourg P, Roze E, Labauge P, Nguyen S. Haematopoietic stem cell transplantation in CSF1R-related adult-onset leukoencephalopathy with axonal spheroids and pigmented glia. *J Neurol Neurosurg Psychiatry*. 2019;90:1375–6. PubMed PMID: 31213485.

- N'Gbo N'Gbo Ikazabo R, Mostosi C, Jissendi P, Labaisse M, Vandernoot I. A new DARS2 mutation discovered in an adult patient. *Case Rep Neurol.* 2020;12:107–13. PubMed PMID: 32308605.
- Parodi L, Coarelli G, Stevanin G, Brice A, Durr A. Hereditary ataxias and paraparesias: clinical and genetic update. *Curr Opin Neurol.* 2018;31:462–71. PubMed PMID: 29847346.
- Scheper GC, van der Klok T, van Andel RJ, van Berkel CG, Sissler M, Smet J, Muravina TI, Serkov SV, Uziel G, Bugiani M, Schiffmann R, Krageloh-Mann I, Smeitink JA, Florentz C, Van CR, Pronk JC, van der Knaap MS. Mitochondrial aspartyl-tRNA synthetase deficiency causes leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation. *Nat Genet.* 2007;39:534–9. PubMed PMID: 17384640.
- Serkov SV, Pronin IN, Bykova OV, Maslova OI, Arutyunov NV, Muravina TI, Kornienko VN, Fadeeva LM, Marks H, Bonnemann C, Schiffmann R, van der Knaap MS. Five patients with a recently described novel leukoencephalopathy with brainstem and spinal cord involvement and elevated lactate. *Neuropediatrics.* 2004;35:1–5. PubMed PMID: 15002045.
- Shimajima K, Higashiguchi T, Kishimoto T, Miyatake S, Miyake N, Takanashi J, Matsumoto N, Yamamoto T. A novel DARS2 mutation in a Japanese patient with leukoencephalopathy with brainstem and spinal cord involvement but no lactate elevation. *Hum Genome Var.* 2017;4:17051. PubMed PMID: 29138691.
- Simons C, Griffin LB, Helman G, Golas G, Pizzino A, Bloom M, Murphy JL, Crawford J, Evans SH, Topper S, Whitehead MT, Schreiber JM, Chapman KA, Tift C, Lu KB, Gamper H, Shigematsu M, Taft RJ, Antonellis A, Hou YM, Vanderver A. Loss-of-function alanyl-tRNA synthetase mutations cause an autosomal-recessive early-onset epileptic encephalopathy with persistent myelination defect. *Am J Hum Genet.* 2015;96:675–81. PubMed PMID: 25817015.
- Steenweg ME, Pouwels PJ, Wolf NI, van Wieringen WN, Barkhof F, van der Knaap MS. Leukoencephalopathy with brainstem and spinal cord involvement and high lactate: quantitative magnetic resonance imaging. *Brain.* 2011;134:3333–41. PubMed PMID: 22006980.
- Steenweg ME, van Berge L, van Berkel CG, de Coo IF, Temple IK, Brockmann K, Mendonça CI, Vojta S, Kolk A, Peck D, Carr L, Uziel G, Feigenbaum A, Blaser S, Scheper GC, van der Knaap MS. Early-onset LBSL: how severe does it get? *Neuropediatrics.* 2012;43:332–8. PubMed PMID: 23065766.
- Stellingwerff MD, Figuccia S, Bellacchio E, Alvarez EK, Castiglioni C, Topaloglu P, Stutterd CA, Erasmus CE, Sanchez-Valle A, Lebon S, Hughes S, Schmitt-Mechelke T, Vasco G, Chow G, Rahikkala E, Dallabona C, Okuma C, Aiello C, Goffrini P, Abbink TEM, Bertini ES, Van der Knaap MS. LBSL: case series and DARS2 variant analysis in early severe forms with unexpected presentations. *Neurol Genet.* 2021;7:e559. PubMed PMID: 33977142.
- Synofzik M, Schicks J, Lindig T, Biskup S, Schmidt T, Hansel J, Lehmann-Horn F, Schöls L. Acetazolamide-responsive exercise-induced episodic ataxia associated with a novel homozygous DARS2 mutation. *J Med Genet.* 2011;48:713–5. PubMed PMID: 21749991.
- Taft RJ, Vanderver A, Leventer RJ, Damiani SA, Simons C, Grimmond SM, Miller D, Schmidt J, Lockhart PJ, Pope K, Ru K, Crawford J, Rosser T, de Coo IF, Juneja M, Verma IC, Prabhakar P, Blaser S, Raiman J, Pouwels PJ, Bevova MR, Abbink TE, van der Knaap MS, Wolf NI. Mutations in DARS cause hypomyelination with brain stem and spinal cord involvement and leg spasticity. *Am J Hum Genet.* 2013;92:774–80. PubMed PMID: 23643384.
- Távora DG, Nakayama M, Gama RL, Alvim TC, Portugal D, Comerlato EA. Leukoencephalopathy with brainstem and spinal cord involvement and high brain lactate: report of three Brazilian patients. *Arq Neuropsiquiatr.* 2007;65:506–11. PubMed PMID: 17665025.
- Uluc K, Baskan O, Yildirim KA, Ozsahin S, Koseoglu M, Isak B, Scheper GC, Gunal DI, van der Knaap MS. Leukoencephalopathy with brain stem and spinal cord involvement and high lactate: a genetically proven case with distinct MRI findings. *J Neurol Sci.* 2008;273:118–22. PubMed PMID: 18619624.

- van Berge L, Hamilton EM, Linnankivi T, Uziel G, Steenweg ME, Isohanni P, Wolf NI, Krägeloh-Mann I, Brautaset NJ, Andrews PI, de Jong BA, al Ghamdi M, van Wieringen WN, Tannous BA, Hulleman E, Würdinger T, van Berkel CG, Polder E, Abbink TE, Struys EA, Scheper GC, van der Knaap MS. Leukoencephalopathy with brainstem and spinal cord involvement and lactate elevation: clinical and genetic characterization and target for therapy. *Brain*. 2014;137:1019–29. PubMed PMID: 24566671.
- van der Knaap MS, van der Voorn P, Barkhof F, van Coster R, Krageloh-Mann I, Feigenbaum A, Blaser S, Vles JS, Rieckmann P, Pouwels PJ. A new leukoencephalopathy with brainstem and spinal cord involvement and high lactate. *Ann Neurol*. 2003;53:252–8. PubMed PMID: 12557294.
- van der Knaap MS, Valk J. *Magnetic Resonance of Myelin Myelination and Myelin Disorders*. 3 ed. Berlin: Springer; 2005.
- van der Knaap MS, Pronk JC, Scheper GC. Vanishing white matter disease. *Lancet Neurol*. 2006;5:413–23. PubMed PMID: 16632312.
- van der Knaap MS, Schiffman R, Mochel F, Wolf NI. Diagnosis, prognosis and treatment of leukodystrophies. *Lancet Neurol*. 2019;18:962–72. PubMed PMID: 31307818.
- Vernon HJ, Bindoff LA. Mitochondrial ataxias. *Handb Clin Neurol*. 2018;155:129–41. PubMed PMID: 29891055.
- Yahia A, Elsayed L, Babai A, Salih MA, Misbah El-Sadig S, Amin M, Koko M, Abubakr R, Idris R, Taha SOMA, Elmalik SA, Brice A, Eltahir Ahmed A, Stevanin G. Intra-familial phenotypic heterogeneity in a Sudanese family with DARS2-related leukoencephalopathy, brainstem and spinal cord involvement and lactate elevation: a case report. *BMC Neurology*. 2018;18:175. PubMed PMID: 30352563.
- Yamashita S, Miyake N, Matsumoto N, Osaka H, Iai M, Aida N, Tanaka Y. Neuropathology of leukoencephalopathy with brainstem and spinal cord involvement and high lactate caused by a homozygous mutation of DARS2. *Brain Dev*. 2013;35:312–6. PubMed PMID: 22677571.
- Yazici Gencdal I, Dincer A, Obuz O, Yapici Z. Leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation (LBSL): a case with long-term follow-up. *Neurologist*. 2020;25:144–7. PubMed PMID: 32925487.
- Yelam A, Nagarajan E, Chuquilin M, Govindarajan R. Leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation: a novel mutation in the DARS2 gene. *BMJ Case Rep*. 2019;12:e227755. PubMed PMID: 30635318.
- Zhang J, Liu M, Zhang Z, Zhou L, Kong W, Jiang Y, Wang J, Xiao J, Wu Y. Genotypic spectrum and natural history of cavitating leukoencephalopathies in childhood. *Pediatr Neurol*. 2019;94:38–47. PubMed PMID: 30770271.

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